



# An integrated platform for drug screening of neurological disease models

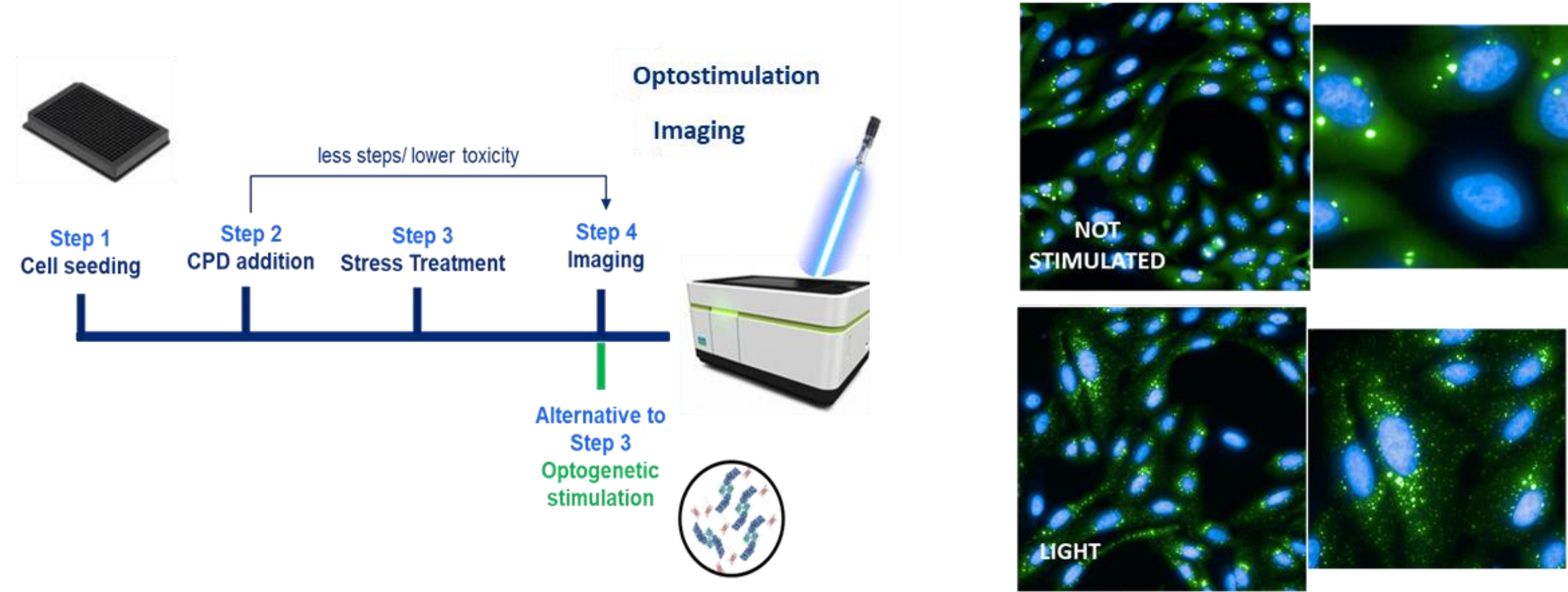
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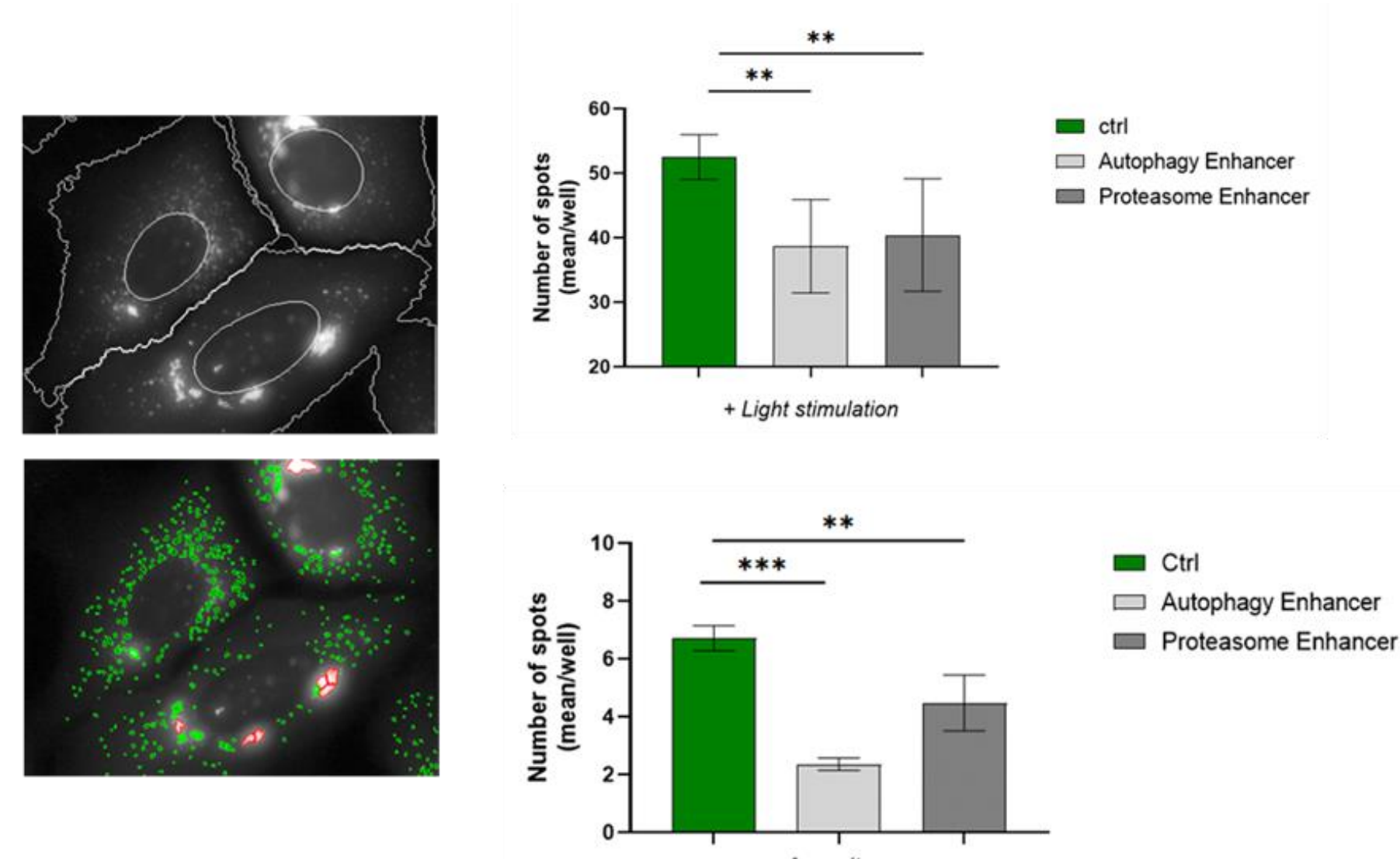
## Introduction

Several human neurodegenerative diseases share a common pathological feature: the formation and accumulation of prion-like aberrant protein aggregates and insoluble inclusions, which lead to progressive neuronal dysfunction and toxicity. Here we present a multilayer platform to address TDP43 dysfunctions, a protein involved in RNA processing and the onset of pathological aggregates, using imaging, optogenetic assays, high-throughput miniaturized formats and relevant cell models, as a proof-of-concept to approach different neuro-related disorders and establishing the basis for therapeutic strategies.

## Biomolecular Condensate/Aggregates Assay

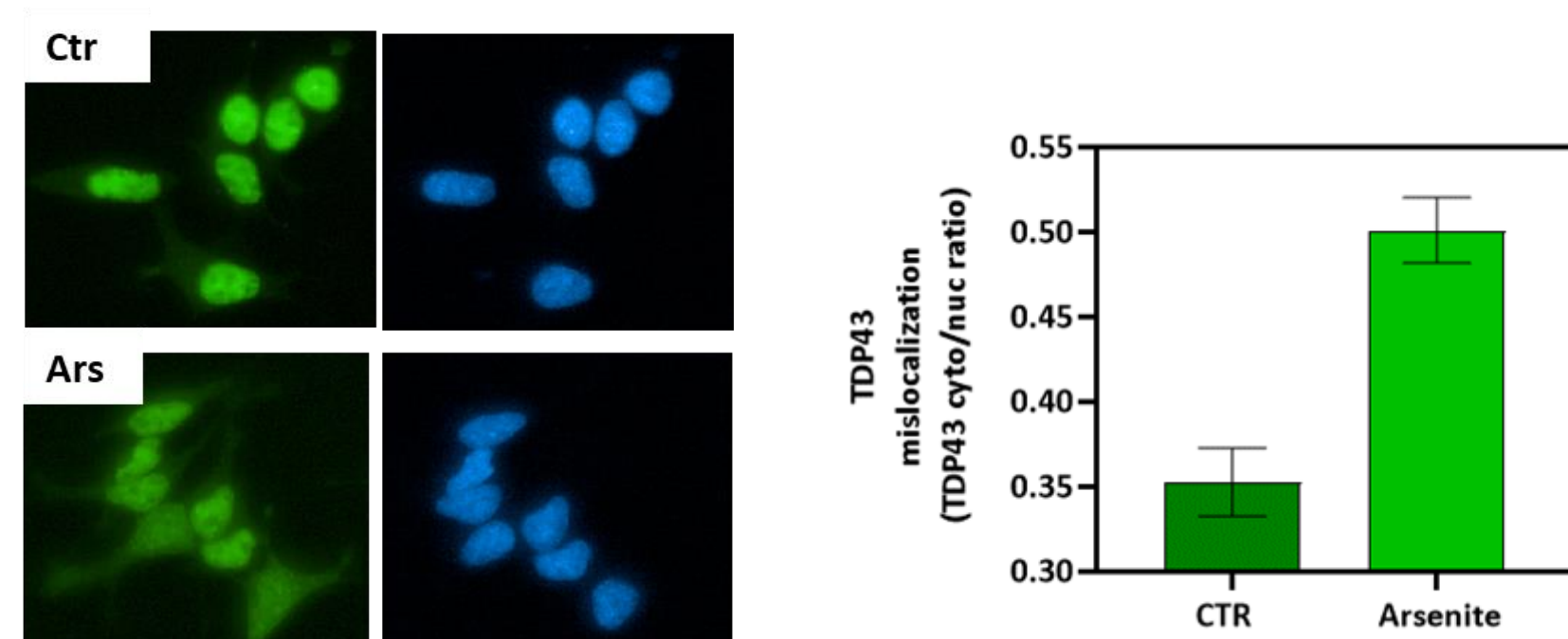


TDP43 biomolecular condensate/aggregation puncta formation by light stimulation (or chemical stress) in U2OS reporter cell lines. Left: Experimental protocol workflow in miniaturized format (384-w). Right: Images pre- and post- light stimulation were acquired by means of a High Content Microscope (Operetta CLS, PerkinElmer); 40X magnification.

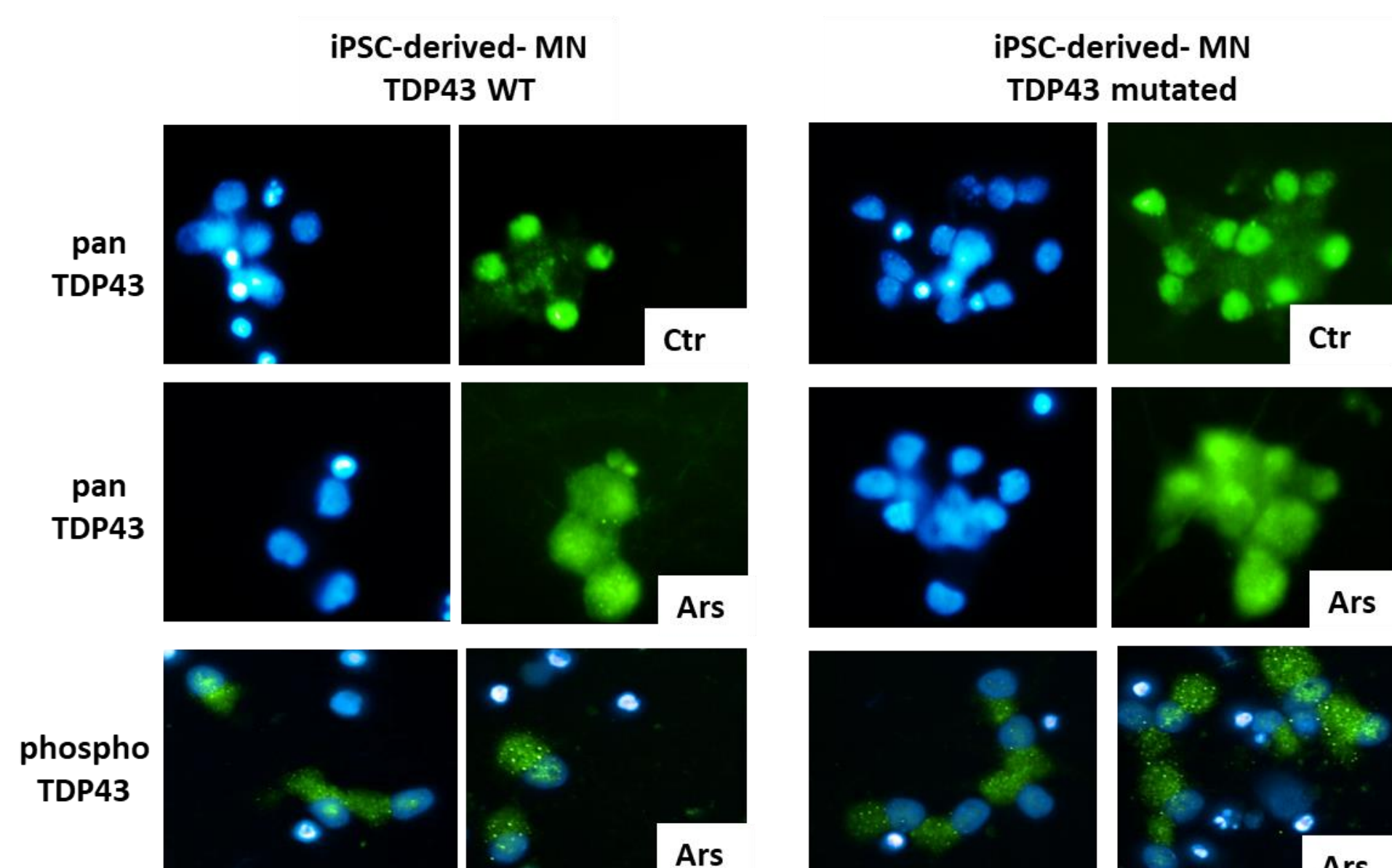


Cellular clearance pathways decrease the TDP43 puncta accumulation. Left: Green objects represent TDP43 positive small condensates, red objects large aggregates. Right: plots of number of spots detected with the two different stimulation protocols (light or chemical stressor, in presence of autophagy or proteasome enhancer drugs).

## TDP43 mis-localization in relevant cell models



TDP43 mis-localization in human neuroblastoma cells. Endogenous level of TDP43 (green channel) upon stressor treatment (Arsenite, Ars) measured by immunostaining in human neuroblastoma cell lines, SH-SY5Y cells, counterstained with Dapi for nuclei (blue channel). Plot represents the ratio of TDP43 cytosol mis-localization.



TDP43 mis-localization and puncta formation in iPSC-derived motor neurons. Analysis of TDP-43 (green channel) localization, upon stressor treatment (Arsenite, Ars), was performed in human iPSC-derived motor neurons (MN; iCell Motor Neurons FCDI) possessing either non-mutant (WT, left panel) or mutant TDP-43 (Mutated, right panel). TDP-43 was measured by means of both total TDP-43 (pan TDP43) and phospho-TDP-43 (phospho TDP43) immunofluorescence and counterstained with DAPI for nuclei (blue channel).

## TDP43

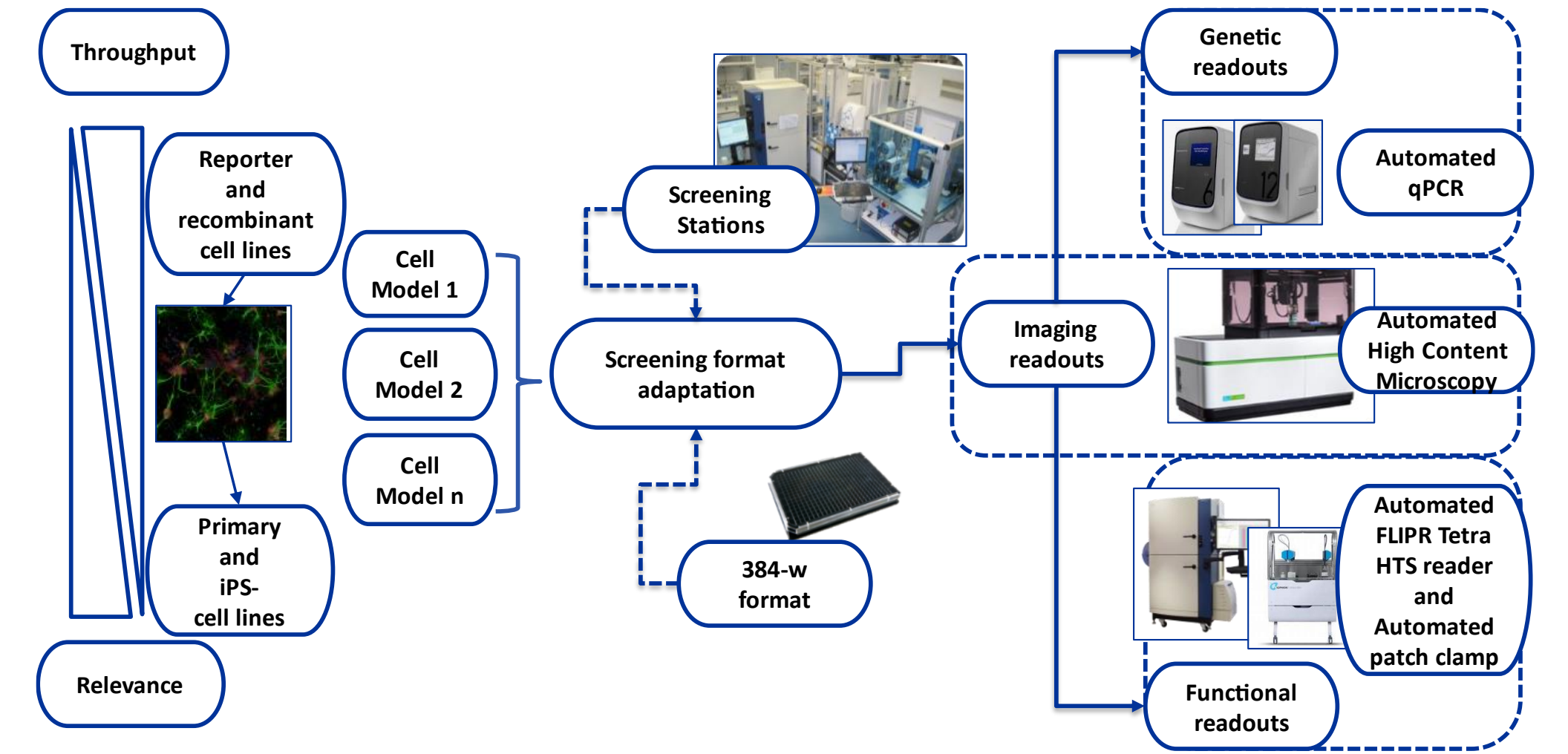


TDP-43 sequence with 2 RNA-recognition motifs (RRMs) and a prion-like, Low Complexity Domain (LCD). LCD is an intrinsically disordered region (IDR) domain, which renders TDP-43 aggregation prone.

LCDs are common in RNA-Binding Proteins (RBPs) and mediate protein-RNA interactions through a liquid-liquid phase separation (LLPS) process.

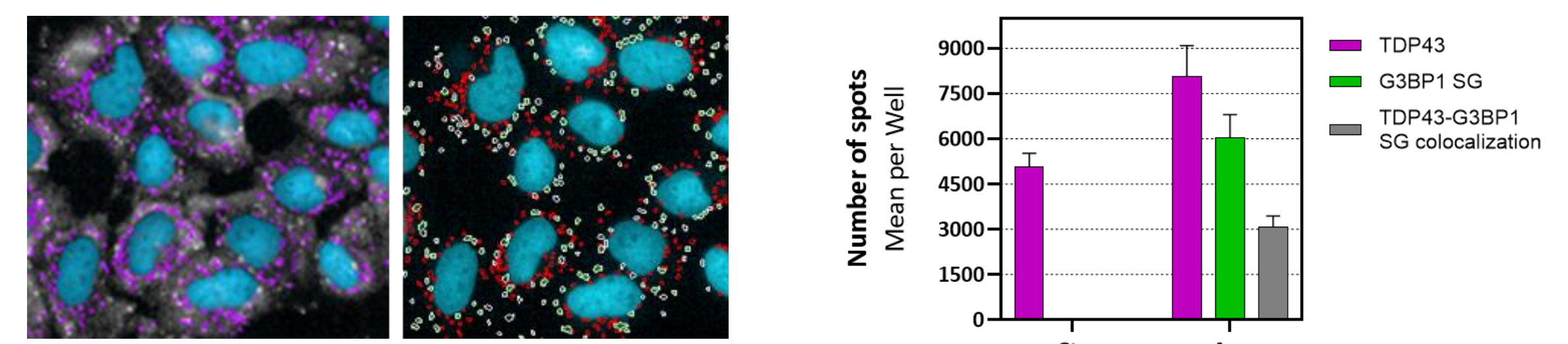
Mutations clustered within the LCD and the RRM domains may lead to several proteinopathies.

## Axxam multilayer framework platform for screening



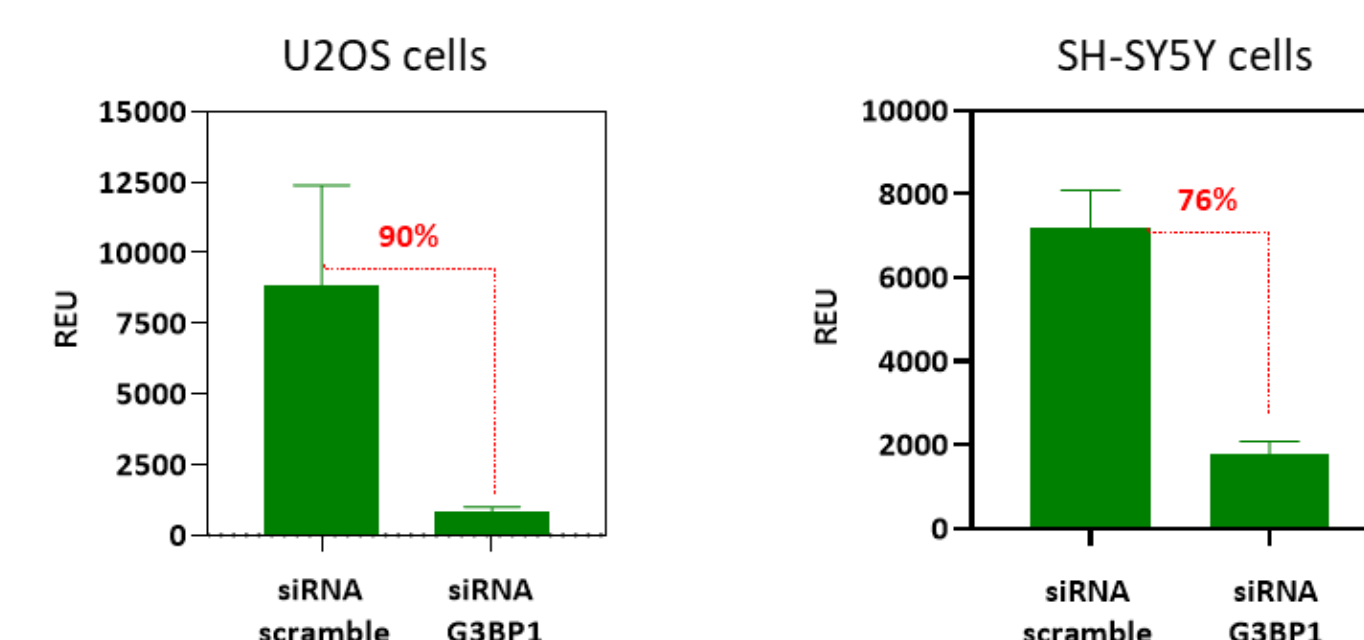
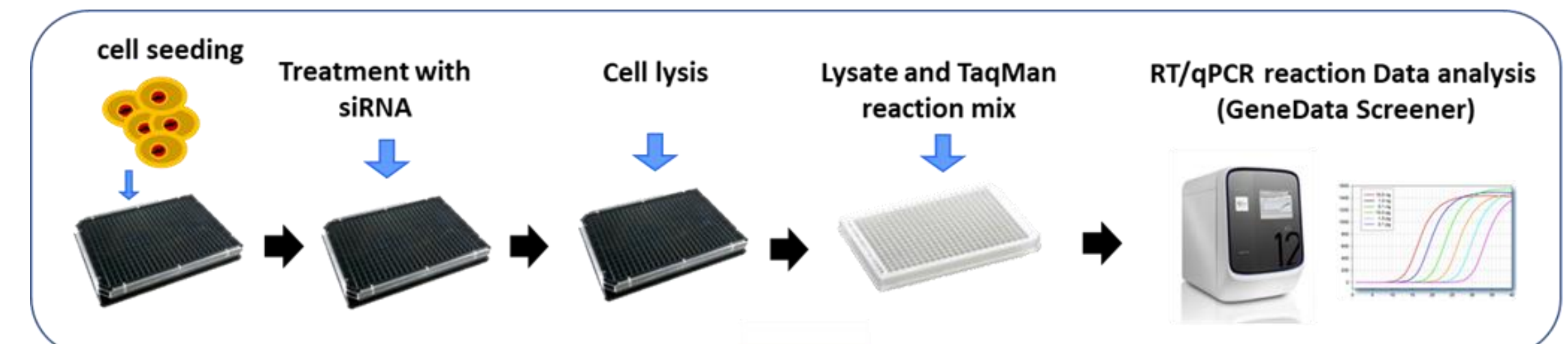
Several assays can be used sequentially and/or in parallel, combining and balancing throughput, relevant models and multiple readouts.

## Stress Granules (G3BP1 positive) Assay

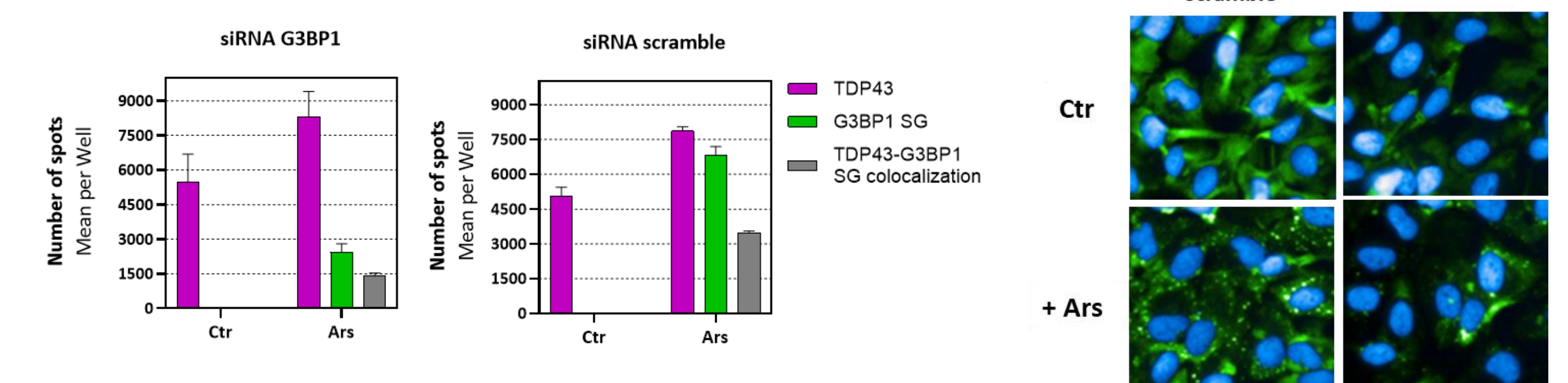


TDP43 and Stress Granules (SG) -G3BP1 positive- co-localization. G3BP1 SG measurement in presence/absence of a stressor treatment (Arsenite, Ars) using multiplexing staining with G3BP1 antibody (green channel), Dapi for nuclei (blue channel) and TDP43 reporter (pink channel). Several features can be analyzed: white spots represent the TDP43 spot co-localized with the G3BP1 positive SG over the discarded red one. Plot: number of puncta in presence and absence of arsenite stressor.

## RNAi automated genomic screening



Stress granule marker G3BP1 efficiently silenced through automated reverse RNAi. Top: High-throughput RT-qPCR workflow in miniaturized (384-well) format and semi-automated conditions. Bottom: G3BP1 mRNA expression levels upon transfection with siRNA G3BP1 pool and siRNA non-targeting control (scramble) pool in two different cell lines. Data presented in Relative Expression Units (REU).



G3BP1 silencing resulted in TDP43 re-localization. Silencing of the SG marker G3BP1 following by arsenite stress (left plots), resulted in both a ~60% reduction of the number of G3BP1 spots and a 40% decrease of TDP43-G3BP1 co-localized puncta, compared to the scramble controls, respectively. A slight increase of cytosolic TDP43 compared to control was also observed after G3BP1 knockdown. Right: Representative images of cells upon silencing in presence/absence of arsenite. Blue channel for Dapi, green channel for G3BP1 staining.

## Conclusion

Here we have shown the use of an integrated platform with several assays structured at different levels of complexity as a proof-of-concept to evaluate the dysfunctions of RNA-binding proteins (RBPs), such as TDP43. The wide-angle approach together with the automated miniaturized formats of the assays can be relevant for the study of neurodegenerative diseases.

References: M. J. et al. *Neuron*, 2019; F. Y. M. et al. *Neuron* 2019; A. S. et al. *Nature Reviews Molecular Cell Biology*, 2021; H. M. et al. *Cell*, 2021

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